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TWO HOUSTO	ON CENTER		ARIANI, KADE	
909 FANNIN, SUITE 3500 HOUSTON, TX 77010			ART UNIT	PAPER NUMBER
			1651	
			NOTIFICATION DATE	DELIVERY MODE
			03/18/2010	ELECTRONIC

Please find below and/or attached an Office communication concerning this application or proceeding.

The time period for reply, if any, is set in the attached communication.

Notice of the Office communication was sent electronically on above-indicated "Notification Date" to the following e-mail address(es):

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Office Action Summary		Application No.	Applicant(s)	Applicant(s)		
		10/544,212	FUKAE, KAZUHII	FUKAE, KAZUHIRO		
		Examiner	Art Unit			
		Kade Ariani	1651			
Period fo	The MAILING DATE of this communicati or Reply	on appears on the cover sheet	with the correspondence a	ddress		
A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE 3 MONTH(S) OR THIRTY (30) DAYS, WHICHEVER IS LONGER, FROM THE MAILING DATE OF THIS COMMUNICATION. - Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication. - If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication. - Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133). Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).						
Status						
•	Responsive to communication(s) filed on This action is FINAL . 2b) Since this application is in condition for a closed in accordance with the practice u	☐ This action is non-final. allowance except for formal ma	•	e merits is		
Dispositi	ion of Claims		,			
5) 6) 7) 8)	Claim(s) <u>1-20</u> is/are pending in the appli 4a) Of the above claim(s) is/are w Claim(s) is/are allowed. Claim(s) <u>1-20</u> is/are rejected. Claim(s) is/are objected to. Claim(s) are subject to restriction in the appli	rithdrawn from consideration.				
10)	The specification is objected to by the ExThe drawing(s) filed on is/are: a)[Applicant may not request that any objection Replacement drawing sheet(s) including the The oath or declaration is objected to by	accepted or b) objected to the drawing(s) be held in abey correction is required if the drawing	rance. See 37 CFR 1.85(a).			
Priority ι	ınder 35 U.S.C. § 119					
 12) Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f). a) All b) Some * c) None of: 1. Certified copies of the priority documents have been received. 2. Certified copies of the priority documents have been received in Application No 3. Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)). * See the attached detailed Office action for a list of the certified copies not received. 						
2) Notic 3) Inform	t(s) se of References Cited (PTO-892) se of Draftsperson's Patent Drawing Review (PTO-9 mation Disclosure Statement(s) (PTO/SB/08) r No(s)/Mail Date	948) Paper N	v Summary (PTO-413) o(s)/Mail Date if Informal Patent Application			

DETAILED ACTION

The amendment filed on December 4,2009, has been received and entered.

Claims 1-20 are pending in this application and were examined on their merits.

Claim Rejections - 35 USC § 103

The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:

(a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negatived by the manner in which the invention was made.

Claims 1-20 are rejected under 35 U.S.C. 103(a) as being unpatentable over Koketsu et al. (J. Carbohydrate Chemistry, 1995, Vol. 14, No.6, p.833-841) and SCORE search results, in view of Yamamoto et al. (JP 08099988 A, 1996, Abstract) and further in view of Inazu et al. (in IDS, Peptide Science 1998, M. Kondo Edition, p. 153-156) and Koketsu et al. (The journal of Food Science, 1993, Vol. 58, No. 4, p.743-747) and Yamamoto, K. (Journal of Bioscience and Bioengineering, 2001, Vol. 92, No. 6, p.493-501).

Claims 1-13 are drawn to a process for preparing asparagine-linked oligosaccharide derivatives comprising the steps of (a) treating a delipidated egg yolk with orientase to obtain a mixture of peptide-linked oligosaccharides (b) treating the

mixture of peptide-linked oligosaccharides with actinase to obtain a mixture of asparagine-linked oligosaccharides, (c) introducing a lipophilic protective group into the asparagine-linked oligosaccharides; and (d) subjecting the mixture of asparagine-linked oligosaccharide derivatives to a fractionating chromatography using a reverse phase column to separate the mixture into individual asparagine-linked oligosaccharide derivatives, delipidated egg yolk is obtained by delipidating an avian egg yolk with an organic solvent, wherein the asparagine-linked oligosaccharide derivatives are undecato penta-saccharide derivatives, wherein the asparagine-linked oligosaccharide derivatives are undeca- to hepta-saccharide derivatives, wherein the asparagine-linked oligosaccharide derivatives are undeca- to nona-saccharide derivatives, asparaginelinked oligosaccharide derivative is an undeca-saccharide derivatives, the lipophilic protective group is a carbonate-containing group, the lipophilic protective group is Fmoc group, wherein the asparagine-linked oligosaccharides contained in the mixture of asparagine-linked oligosaccharide derivatives obtained by step (b) are hydrolyzed before the subsequent step to cut off some sugar moieties, wherein the asparaginelinked oligosaccharides contained in the mixture of asparagine-linked oligosaccharide derivatives obtained in the mixture by step (c) are hydrolyzed before the subsequent step to cut off some sugar moieties, and wherein the asparagine-linked oligosaccharide derivatives have the following formula (claim 13), wherein Prot is a lipophilic protective group, Asn is an asparagine, and Ac is an acetyl group.

Claims 14-20 are drawn a to a process for preparing asparagine-linked oligosaccharide derivatives comprising the steps of (a) treating a delipidated egg yolk

with a protease (to obtain a mixture of peptide-linked oligosaccharides); (b) isolating the mixture of peptide-linked oligosaccharides; (c) treating the mixture of peptide-linked oligosaccharides with a peptidase (to obtain a mixture of asparagine-linked oligosaccharides); (d) introducing a lipophilic protective group into the asparagine-linked oligosaccharides in the mixture (to obtain a mixture of asparagine-linked oligosaccharides derivatives), the process further comprising (e) subjecting the mixture of asparagine-linked oligosaccharide derivatives to a fractionating chromatography using a reverse phase column (to separate the mixture into individual asparagine-linked oligosaccharide derivatives), wherein asparagine-linked undeca-saccharide derivatives, and asparagine-linked undeca-saccharide derivatives have the formula of claim 20.

Koketsu et al. (1995) teach a process for preparing asparagine-linked oligosaccharide derivatives comprising, treating a delipidated egg yolk with orientase (a protease) to obtain a mixture of peptide-linked oligosaccharides, isolating the mixture of peptide-linked oligosaccharides, an undeca-saccharide derivatives, and the asparagine-linked oligosaccharide derivatives structural formula (p.838 3rd paragraph lines 1-5, and Figure 3., see the SCORE structure search result). Koketsu et al. also teach purification by fractionating chromatography (Abstract and p.839 2nd paragraph).

Koketsu et al. do not teach treating the mixture of peptide-linked oligosaccharides with actinase, introducing a lipophilic protective group into the asparagine-linked oligosaccharides in the mixture, the lipophilic protective group is Fmoc group, and wherein the asparagine-linked oligosaccharide derivatives obtained are hydrolyzed before the subsequent step to cut off some sugar moieties. However,

Yamamoto et al. (1996) teach treating delipidated egg yolk with actinase E (a peptidase), to enzymatically digest the insoluble proteins obtained form delipidated (defatted) egg yolk and extract oligosaccharides (Abstract).

Inazu et al. teach a process for preparing asparagine-linked oligosaccharide derivatives, introducing a lipophilic protective group to the asparagine-linked oligosaccharides, and subjecting the mixture of asparagine-linked oligosaccharide derivatives to a fractionating chromatography using a reverse phase column to separate the mixture (p. 153, Abstract and p. 154, Figure 1. step 1, and p.156 1st paragraph lines 4-5).

Koketsu et al. (1993) teach egg yolk can be delipidated by treating with ethanol (organic solvent), and separating the mixture of oligosaccharides by reverse-phase column. Koketsu et al. also teach oligosaccharide derivatives can be obtained by hydrolyzing (cut off some sugar moieties), and obtaining an undeca saccharide derivative (Abstract, p.743 2nd column, 3rd paragraph, lines 1-2, p. 744, 2nd column 4th paragraph, lines 1-5, and p. 746, Figure 5, 3rd oligosaccharide derivative). Koketsu et al. further teach sialyloligosaccharides are being used to create drugs and food companies formulate functional foods by addition of sialyloligosaccharides. Koketsu et al. teach chemical methods for preparation of sialyloligosaccharides are cumbersome and laborious (p.743 Introduction, 1st column 2nd paragraph).

Further motivation is in Yamamoto (2001) who teaches glycosylated peptide containing asparagine-linked oligosaccharide (N-acetylglucosaminyl peptide with an N-acetylglucosamine moiety bound to the asparaginyl residue of the peptide) have higher

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degree of resistance to protease digestion (Abstract). Yamamoto further teaches chemical synthesis of oligosaccharides are labor–intensive and involve complicated steps, on the other hand, enzymatic methods have the advantages because of their high stereo- and regio-selectivities (p. 493 Introduction 1st column, 1st paragraph).

Therefore, in view of the above teachings, a person of ordinary skill in the art at the time the invention was made, knowing that asparagine-linked oligosaccharide have higher degree of resistance to protease digestion and that chemical synthesis of oligosaccharides are labor-intensive and the advantages of enzymatic methods, would have been motivated to modify the process as taught by Koketsu et al. by using the actinase (peptidase) according to the teachings of Yamamoto et al. and by introducing a lipophilic protective group (Fmoc) into the asparagine-linked oligosaccharide to hydrolyze the peptide-linked oligosaccharide according to teach teachings of Inazu et al. and Koketsu et al. (1993), in order to provide a process for preparing asparagine-linked oligosaccharide derivatives with a reasonable expectation of success, because Yamamoto et al. teach treating delipidated egg yolk with actinase E to extract useful oligosaccharides from delipidated egg yolk, because Inazu et al. teach lipophilic protective group (Fmoc) can be introduced to asparagine-linked oligosaccharide to ease the purification, and because Koketsu et al. teach forming asparagine-linked oligosaccharide derivatives by hydrolyzing sugar moieties.

Answer to Arguments

Applicant's arguments with respect to claims 1-20 filed on 12/4/2009 have been considered but are fully considered but are not persuasive.

Applicant argues that Yamamoto(1996) teaches away from directly treating the delipidated egg yolk with enzymes, this argument is not found persuasive because Yamamoto et al. teaching does not criticize, discredit, or otherwise discourage the solution claimed, since Yamamoto et al. (1996) teach using actinase E, to enzymatically digest the insoluble proteins obtained form delipidated (defatted) egg yolk and extract oligosaccharides.

In response to applicant's argument that the examiner's conclusion of obviousness is based upon improper hindsight reasoning, it must be recognized that any judgment on obviousness is in a sense necessarily a reconstruction based upon hindsight reasoning. But so long as it takes into account only knowledge which was within the level of ordinary skill at the time the claimed invention was made, and does not include knowledge gleaned only from the applicant's disclosure, such a reconstruction is proper. See *In re McLaughlin*, 443 F.2d 1392, 170 USPQ 209 (CCPA 1971).

In this case, as mentioned immediately above, Yamamoto (2001) teaches a glycosylated peptide containing asparagine-linked oligosaccharide have higher degree of resistance to protease digestion (Abstract). Yamamoto further teaches chemical synthesis of oligosaccharides are labor–intensive and involve complicated steps, on the other hand, enzymatic methods have the advantages because of their high stereo- and regio-selectivities (p. 493 Introduction 1st column, 1st paragraph). Therefore, a person of

ordinary skill in the art at the time the invention was made would have realized that glycosylated peptides containing asparagine-linked oligosaccharide have higher degree of resistance to protease digestion and more specific peptidases are required to liberate these peptides.

Moreover, Inazu et al. teach a process for preparing asparagine-linked oligosaccharide derivatives, introducing a lipophilic protective group to the asparagine-linked oligosaccharides, and subjecting the mixture of asparagine-linked oligosaccharide derivatives to a fractionating chromatography using a reverse phase column to separate the mixture (p. 153, Abstract and p. 154, Figure 1. step 1, and p.156 1st paragraph lines 4-5).

Koketsu et al. (1993) teach egg yolk can be delipidated by treating with ethanol (organic solvent), and separating the mixture of oligosaccharides by reverse-phase column. Koketsu et al. also teach oligosaccharide derivatives can be obtained by hydrolyzing (cut off some sugar moieties), and obtaining an undeca saccharide derivative (Abstract, p.743 2nd column, 3rd paragraph, lines 1-2, p. 744, 2nd column 4th paragraph, lines 1-5, and p. 746, Figure 5, 3rd oligosaccharide derivative). Koketsu et al. further teach sialyloligosaccharides are being used to create drugs and food companies formulate functional foods by addition of sialyloligosaccharides. Koketsu et al. teach chemical methods for preparation of sialyloligosaccharides are cumbersome and laborious (p.743 Introduction, 1st column 2nd paragraph).

Therefore, in view of the above teachings, a person of ordinary skill in the art at the time the invention was made, knowing that asparagine-linked oligosaccharide have

higher degree of resistance to protease digestion and that chemical synthesis of oligosaccharides are labor–intensive and the advantages of enzymatic methods, would have been motivated to modify the process as taught by Koketsu et al. by using the actinase (peptidase) according to the teachings of Yamamoto et al. and by introducing a lipophilic protective group (Fmoc) into the asparagine-linked oligosaccharide to hydrolyze the peptide-linked oligosaccharide according to teach teachings of Inazu et al. and Koketsu et al. (1993), in order to provide a process for preparing asparagine-linked oligosaccharide derivatives with a reasonable expectation of success, because Yamamoto et al. teach treating delipidated egg yolk with actinase E to extract useful oligosaccharides from delipidated egg yolk, because Inazu et al. teach lipophilic protective group (Fmoc) can be introduced to asparagine-linked oligosaccharide to ease the purification, and because Koketsu et al. teach forming asparagine-linked oligosaccharide derivatives by hydrolyzing sugar moieties.

Conclusion

No claims are allowed.

THIS ACTION IS MADE FINAL. See MPEP § 706.07(a). Applicant is reminded of the extension of time policy as set forth in 37 CFR 1.136(a).

A shortened statutory period for reply to this final action is set to expire THREE MONTHS from the mailing date of this action. In the event a first reply is filed within TWO MONTHS of the mailing date of this final action and the advisory action is not

mailed until after the end of the THREE-MONTH shortened statutory period, then the shortened statutory period will expire on the date the advisory action is mailed, and any extension fee pursuant to 37 CFR 1.136(a) will be calculated from the mailing date of the advisory action. In no event, however, will the statutory period for reply expire later than SIX MONTHS from the date of this final action.

Any inquiry concerning this communication or earlier communications from the examiner should be directed to Kade Ariani whose telephone number is (571) 272-6083. The examiner can normally be reached on IFP.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Michael Wityshyn can be reached on (571) 272-0926. The fax phone number for the organization where this application or proceeding is assigned is 571-273-8300.

Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see http://pair-direct.uspto.gov. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free). If you would like assistance from a USPTO Customer Service Representative or access to the automated information system, call 800-786-9199 (IN USA OR CANADA) or 571-272-1000.

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Kade Ariani Examiner Art Unit 1651 /Leon B Lankford/ Primary Examiner, Art Unit 1651